THE UNDERLYING BENEFITS OF IMPROVED FARM PERFORMANCE USING NOVAQPRO[®] MICROBIAL BIOMASS FOR TIGER SHRIMP Penaeus monodon

- Simon C¹, Noble TH¹, Mendoza Porras O¹, Rusu A¹, Rao M¹, Bourne N¹, Hines B¹, Truong HH¹, Rombenso A¹ Charles T², West M², Briggs M³, Smullen R³ and Wade NM^{1,4*}
- 1. CSIRO Agriculture and Food, Brisbane, Australia; 2. Australian Prawn Farms Pty Ltd, Ilbilbie, Queensland, Australia; 3. Ridley Agriproducts Pty Ltd, Narangba Queensland, Australia.4. The Roslin Institute, University of Edinburgh, Scotland, UK

INTEGRATED SUSTAINABLE AQUACULTURE PRODUCTION www.csiro.au

- Balancing the competing demands of increased feed costs and reduced farm gate prices with lowering marine capture resources and nutrient inputs is critical to ensure the long-term profitability and sustainability of shrimp farming.
- The use of least-cost formulations may seem attractive, but it often negatively impacts animal health and performance, reducing yields, water quality, and nutritional condition. Poor feed utilisation in turn affects water quality and lead to excessive nutrient emissions from the farm.
- The inclusion of a microbial biomass commercialised as NovaqPro[®] has enabled greater flexibility in formulations, for instance the reduction of protein and marine

resources in the diet of shrimp^{1,2}, and improving survival through disease resistance and immune stimulation^{3,4}.

To characterise the benefits at the commercial scale, a replicated pond experiment was conducted at Australian Prawn Farms in Australia



- Full commercial grow out cycle over 190-200 days to harvest •
- Eco or Propel-G (contains NovaqPro[®]) fed from day 70
- Statistics: t-tests (P<0.05)









Improved survival, pond yield and feed conversion

	ECO	PROPEL-G	% GAIN	Statistics
Average Harvest Weight (g)	30.0 ± 0.8	31.4 ± 0.8	4.7	nsd
Average Survival (%)	80.2 ± 5.2	88.2 ± 5.0	10.0	nsd
Average Pond Yield (kg)	16809 ± 1085	19320 ± 728	14.9	nsd
Average FCR	2.02 ± 0.04	1.89 ± 0.03	-6.1	P = 0.04
Total Ammonia Nitrogen (mg/L)	1.42 ± 0.22	1.00 ± 0.19	-29.0	nsd
BRIX (nutritional condition)	16.0 ± 0.8	17.7 ± 0.4	10.8	nsd

CSIRO

Reduced pond dissolved total ammonia nitrogen levels



100-110 110-120 120-130 130-140

NUTRIENT RETENTION ANALYSIS N and P retention, 4 ponds/diet DOC 70-130 and DOC 70-harvest

HEMOLYMPH ANALYSIS BRIX refractometry DOC150 4 ponds/diet, 28-36 individuals/pond

6500 QTRAP MS), 50 peptides targeting 26 functional proteins^{5,} 4 ponds/diet, 5 pools/pond of 6-8 individuals



Species diversity (number and evenness) affected by diet and reduced towards end of crop





Targeted quantification of 27 functional proteins:



100% 90% 70% 60% 50% 40%

Days of Culture (DOC)

Increased proportion of large sized prawns at harvest Size Splits Eco vs Propel-G



NovagPRo

Propel-G enabled: ~5% greater average final weight ~10% increased survival Reduced FCR by 6% ~40% reduction in TAN

~15% increased harvest

Tissue used - Intestine without digesta, frozen & RNALater preserved

MICROBIOME ANALYSIS

16s V123 sequencing

3 ponds/diet, 19 individuals/pond

OC 70 (initial), DOC 125, DOC 150



References:

Actibacte Actibacteriu

Blastopirellu

- 1. Simon et al (2020) Aquaculture. 520. 734679
- 2. Glencross et al (2014). Aquaculture, 431, 12-19.
- 3. Sellars et al (2015) Journal of the World Aquaculture Society. 46, 328-336.
- 4. Noble et al (2023) Animal Feed Science and Technology. 299, 115626.
- 5. Mendoza-Porras et al (2023) Comp Biochem Phys D. 46.



FOR FURTHER INFORMATION

Cedric Simon e cedric.simon@csiro.au Nick Wade nwade@ed.ac.uk е



ğ

2 TA







ACKNOWLEDGEMENTS

This study was funded through a collaborative research alliance between CSIRO and Ridley Corp. Australian Prawn Farms (APF): Andrew Smith **Ridley:** Gustavo Lehnebach