



## for the detection of jellyfish in aquaculture system

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### ABSTRACT

Scyphozoa, commonly referred to as jellyfish within the Cnidaria, can lead mass mortality in aquaculture and fisheries through a direct harmful impact with their toxins derived from their nematocysts or indirect effects such as gill malfunctions. In addition, their adhesive polyps can directly attach to aquaculture systems and produce ephyra and medusae. Jellyfish blooms are becoming more frequent due to global warming and increased anthropogenic activity, which means their impact on an aquaculture and fisheries will become increasingly significant. Efficient and precise monitoring of jellyfish occurrences and blooms is important to prevent subsequent problems and find solutions. Therefore, we designed new COI target primer sets with higher resolution for the detection of jellyfish and conducted an *in silico* evaluation. Scyphozoa COI sequences were assembled from NCBI search results for "Scyphozoa, complete" and aligned with MEGA-X software. Primers were designed to cover highly variable regions, meeting criteria such as GC%, amplicon size, and primer length. Consequently, primer sets were developed to generate approximately 350 bp amplicons. The accuracy of the newly designed primers was evaluated using BLASTn and PrimerMiner in R package. With these detection methods, we expect rapid detection of jellyfish bloom both in and outside of aquaculture environments.

### Method

1. Sequence collect

2. Alignment

3. Primer design

4. *In silico* PCR

5. *In vitro* validation



- Synthesized gBlock DNA
- PCR

### Results

Fig 1 Location within COI sequence of newly designed primer set



Fig 2 Sequence logos represent the base of Scyphozoa targeted primer regions and below show the designed primer sequences.

> Forward



> Reverse

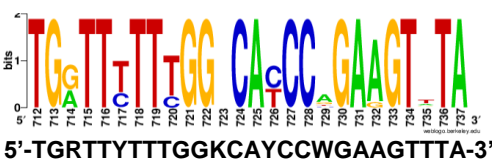


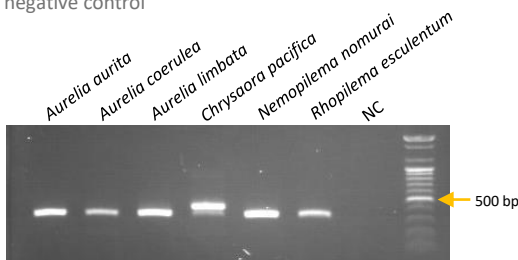
Fig 3 Comparison of the *in silico* PCR results of existing primers and newly designed primer set

Threshold value was specified as 200.

Species name (genbank number)	<i>In silico</i> PCR result	
	Universal	Designed
Acromitus flagellatus (NC_061659)	Fail	OK
Catostylus townsend (NC_061766)	Fail	Fail
Cassiopea xamachana (NC_016466)	Fail	Fail
Cassiopea sp. MKL-2023 (OR400206)	Fail	OK
Phyllorhiza punctata (OR400204)	Fail	OK
Phyllorhiza punctata (NC_084193)	Fail	OK
Mastigias papus (OQ695499)	Fail	OK
Stomolophus sp. CG-2019 (MK157198)	Fail	Fail
Rhopilema esculentum (KY454768)	Fail	Fail
Nemopilema nomurai (NC_035740)	Fail	OK
Aurelia aurita (NC_008446)	Fail	Fail
Aurelia limbata (NC_046691)	Fail	OK
Aurelia aurita (HQ694729)	Fail	OK
Aurelia coerulea (MT023105)	Fail	OK
Aurelia sp. 4 (LC005414)	Fail	OK
Aurelia sp. 3 (LC005413)	Fail	OK
Pelagia noctiluca (NC_080358)	Fail	OK
Chrysaora quinquecirrha (MW401676)	Fail	OK
Chrysaora pacifica (NC_046775)	Fail	OK
Chrysaora quinquecirrha (HQ694730)	Fail	OK

Fig 4 Electrophoresis result of gBlock DNA amplification through PCR method

NC; negative control



### Conclusion

- ✓ The newly designed primer set is expected to be able to rapid detection of jellyfish bloom both in and outside of aquaculture environments.
- ✓ Easily detecting and responding quickly to polyp stages that are difficult to identify will help solve problems such as increasing weight of nets, causing deformation, and reducing buoyancy of the structures that easily occur in aquaculture.
- ✓ Tests on actual DNA sample obtained from environmental are required to confirm applicability in the field.